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FLUORINATION OF PROTEINS AND PEPTIDES
FOR F-18 POSITRON EMISSION TOMOGRAPHY

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BACKGROUND OF THE INVENTION

5 Field of the Invention:

The present invention concerns methods for radiolabeling proteins and peptides with fluorine-18 (F-18). More particularly, these proteins and peptides are radiolabeled with F-18 by reacting a thiol group contained therein with an F-18-bound labeling reagent which also has a group that is reactive with thiols. The resulting F-18-labeled proteins and peptides are useful in imaging targeted tissue by clinical positron emission tomography.

Description of the Related Art:

Positron emission tomography (PET) is a high resolution, non-invasive, imaging technique for the visualization of human disease. In PET, 511 keV gamma photons produced during positron annihilation decay are detected. In the clinical setting, fluorine-18 (F-18) is one of the most widely used positron-emitting nuclides. F-18 has a half-life ($t_{1/2}$) of 110 minutes, and emits β^+ particles at an energy of 635 keV. It is 97% abundant.

The short half-life of F-18 has limited or precluded its use with longer-lived specific targeting vectors such as antibodies, antibody fragments, recombinant antibody constructs and longer-lived receptor-targeted peptides. In addition, complicated chemistry has been required to link the inorganic fluoride species to such organic targeting vectors. In typical synthesis methods, an intermediate is radiofluorinated, and the F-18-labeled intermediate is purified for coupling to protein amino groups. See, e.g., Lang *et al.*, *Appl. Radiat. Isol.*, 45 (12): 1155-63 (1994); Vaidyanathan *et al.*, *Bioconj. Chem.*, 5: 352-56 (1994).

These methods are tedious to perform and require the efforts of specialized professional chemists. They are not amenable to kit formulations for use in a clinical setting. Multiple purifications of intermediates are commonly required, and the final step, involving linkage to protein lysine residues, usually results in 30-60 % yields, necessitating a further purification step prior to patient administration. In addition, these methods result in fluorinated targeting species which accumulate in the kidney, somewhat like radiometals.

It was recently reported that ^{18}F -fluoroiodomethane ($^{18}\text{FCH}_2\text{I}$) is a useful intermediate for the fluorination of organic intermediates. Zheng *et al.*, *J. Nucl. Med.*, 38: 177P (Abs. 761) (1997). In this process, diiodomethane is fluorinated with the F-18 ion by a room temperature reaction in acetonitrile solvent, resulting in up to a 40% yield. The $^{18}\text{FCH}_2\text{I}$ is then distilled into reaction vials containing various strong nucleophiles in anhydrous acetonitrile and allowed to react at 80°C for fifteen minutes. Nucleophilic attack by carboxylates, thiolates, phenolates, and amines in particular, replaces the remaining iodine of $^{18}\text{FCH}_2\text{I}$, with overall yields of 10 to 35 %. The reaction products can be purified by reverse-phase HPLC. Fluoromethyl diethylamine, fluoromethyl benzoate, fluoromethyl benzyl thioether and fluoromethyl 4-(2-hydroxy-3-aminopropoxy)-carbazole have been made by this method.

As discussed above, the currently available methods for labelling protein-based targeting vectors with F-18 are unsuitable. There is a need, therefore, for a simple, efficient method for incorporating the F-18 radionuclide into peptide-containing targeting vectors, such as proteins, antibodies, antibody fragments, and receptor-targeted peptides, to allow the use of such targeting vectors in routine clinical positron emission tomography.

SUMMARY OF THE INVENTION

The present invention provides methods for incorporating the F-18 radionuclide into peptide-containing targeting vectors.

In accordance with one embodiment of the invention there is provided a method for radiolabeling thiol-containing peptides with fluorine-18 (F-18), comprising reacting a peptide comprising a free thiol group with a labelling reagent having the general formula $^{18}\text{F}-(\text{CH}_2)_m-\text{CR}_1\text{R}_2-(\text{CH}_2)_n-\text{X}$, wherein:

n is 0, 1 or 2;

m is 0, 1 or 2;

and $n+m$ is 0, 1, or 2;

X is selected from the group consisting of iodide, bromide, chloride, azide,
5 tosylate, mesylate, nosylate, triflate, unsubstituted maleimide, maleimide substituted
with one or two alkyl groups, and maleimide substituted with a sulfonate group; and

R_1 and R_2 are the same or different and are selected from the group consisting of
iodide, bromide, chloride, azide, tosylate, mesylate, nosylate, triflate, hydrogen,
-CONH₂, carboxyl, hydroxyl, sulfonic acid, tertiary amine, quaternary ammonium,
10 unsubstituted alkyl, substituted alkyl, -COOR', -CONR'₂, or COR', wherein the
substituents of the substituted alkyl groups are selected from the group consisting of
-CONH₂, carboxyl, hydroxyl, sulfonic acid, tertiary amine and quaternary ammonium
and wherein R' is a C₁-C₆ alkyl or phenyl.

In accordance with another embodiment, there is provided a method for
15 radiolabeling thiol-containing peptides with F-18, comprising reacting a peptide
comprising a free thiol group with a F-18 fluorinated alkene, wherein at least one of the
two double-bonded carbon atoms bears at least one leaving group selected from the
group consisting of iodide, bromide, chloride, azide, tosylate, mesylate, nosylate and
triflate.

20 In accordance with another embodiment of the invention, a peptide that has
been radiolabeled with F-18 as described above is delivered to a targeted tissue
using a bispecific antibody (bsMAb) or a bispecific antibody fragment (bsFab)
containing at least one arm that is specific to the targeted tissue and at least one
other arm that is specific to the F-18-labeled peptide or a low molecular weight
25 hapten conjugated to the F-18-labeled peptide.

In this methodology, the bsMAb or the bsFab is administered to a patient
and allowed to localize to the targeted tissue. Some time later (after the unbound
bsMAb or the unbound bsFab is allowed to clear), the F-18-labeled peptide or the
hapten conjugate thereof is administered to the patient. Since at least one of the
30 arms of the bsMAb or the bsFab is specific to the F-18-labeled peptide or the
hapten conjugated to the F-18-labeled peptide, the F-18-labeled peptide is also
localized to the target. After the unbound F-18-labeled peptide or the unbound

hapten conjugate thereof is allowed to clear, the target is then visualized by routine clinical positron emission tomography.

5 The bsMAb or bsFab is ideally monoclonal and humanized. Preferably, the F-18-labeled peptide contains a thiol group. Examples of suitable peptides are X-Gly-D-Tyr-D-Trp-Gly-D-Lys(X)-Gly-D-Tyr-D-Trp-OH wherein X represents a free or protected amino acid group, Ac-Cys(Y)-D-Tyr-D-Trp-Gly-D-Cys(Y)-Gly-D-Tyr-D-Trp-OH wherein Y represents a free or protected thiol group, and Ac-Gly-D-iodo-Tyr-D-Trp-Gly-D-Lys(Ac)-Gly-D-iodo-Tyr-D-Trp-OH. The hapten can be a metal chelate complex comprising, for example, manganese, iron, or gadolinium
10 which are useful in magnetic resonance imaging (MRI).

The bsMAb, bsFab, and associated methodologies described above are disclosed in U.S. Provisional Application Serial No. 60/090,142 (entitled "Production and use of novel peptide-based agents for use with bispecific antibodies" and filed June 22, 1998), the entire contents of which are herein
15 incorporated by reference.

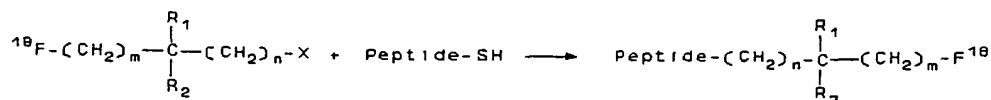
These and other objects and aspects of the invention will become apparent to the skilled artisan in view of the teachings contained herein.

DETAILED DESCRIPTION OF THE INVENTION

20 The present invention provides simple and efficient methods for incorporating the F-18 radionuclide into peptide-containing targeting vectors, such as proteins, antibodies, antibody fragments and receptor-targeted peptides. For convenience, the term "peptide" is used below and in the claims to refer to proteins, antibodies, antibody fragments and receptor-targeted peptides. The methods of the present invention makes such targeting vectors available for routine clinical positron emission tomography.

25 Of all nucleophiles present on peptides, only the free thiol group can be rapidly alkylated at neutral pH and moderate temperature. The present invention takes advantage of this unique property of free thiol groups, and provides methods for labelling thiol-containing peptides with F-18.

In accordance with one embodiment, the method of the present invention
30 comprises the following reaction:



wherein n is 0, 1 or 2, m is 0, 1 or 2, and n+m is 0, 1, or 2, and X is a substitutable leaving group such as iodide, bromide, chloride, azide, tosylate, mesylate, nosylate, triflate and the like. Alternatively, X is maleimide or a substituted maleimide, substituted, for example with one or two alkyl groups or a sulfonate group. Examples of suitable substituted maleimides include 3-methylmaleimide, 3,4-dimethylmaleimide and 3-sulfo-maleimide. R₁ and R₂ can be the same or different and, as discussed in more detail below, are chosen for the desirable physical properties they bring to the reagent. In general, R₁ and R₂ can be selected from the same groups as X, and can be the same as or different from X. Alternatively, R₁ and R₂ each independently can be hydrogen, a substituted or unsubstituted linear or branched alkyl group, or a carbonyl function such as an ester, amide or ketone, for example, -COOR', -CONR'₂, or COR', where R' is a C₁-C₆ alkyl or phenyl. Examples of suitable R₁ and R₂ groups or substituents thereon also include groups which impart aqueous solubility, such as -CONH₂, carboxyl, hydroxyl, sulfonic acid and tertiary amine or quaternary ammonium.

In accordance with another embodiment of the invention, the peptide is labeled with an F-18 fluorinated alkene, wherein at least one of the two double-bonded carbon atoms bears at least one leaving group selected from the group consisting of iodide, bromide, chloride, azide, tosylate, mesylate, nosylate and triflate. Examples of suitable fluorinated alkenes include ¹⁸F-CH=CI₂, ¹⁸F-CI=CH₂, or ¹⁸F-CI=CI₂. The labeling reaction is analogous to the one described above.

The methods of present invention can be used to label any thiol-containing peptide. Of particular interest are peptides useful as targeting vectors. Examples of such targeting vectors include antibodies, F(ab')₂, F(ab)₂, Fab' and Fab fragments, single-chain sub-fragments such as sFvs, divalent constructs such as dsFvs, and polypeptides containing one or more free thiol groups. See Choi *et al.*, *Cancer Res.*, 55: 5323-29 (1995). Further examples include antibody constructs such as antibodies

comprising IgG₃ or IgG₃-F(ab')₂ frameworks. IgG₃'s have multiple hinge-region disulfide groups which can be reduced to generate multiple free thiol groups.

Peptides that originally do not comprise a free thiol group can be labelled in accordance with the present invention by first modifying the peptide to add a free thiol group by methods known to those skilled in the art. For example, the peptide can be thiolated with reagents such as 2-iminothiolane, or intrinsic disulfide bonds such as cystine residues can be reduced. A combination of both modifications also can be performed, such as the acylation of lysine residues with N-succinimidyl-3-(2-pyridylthio)-propionate (SPDP) followed by the controlled reduction of the appended disulfide bond.

In one embodiment of the present invention, the peptide is a Fab or Fab' fragment. These peptides have free thiol groups in their hinge-region, a site which is both specific and remote from the antigen-targeting sites.

To optimize the reaction with the thiol-containing peptides, the labelling reagent preferably has the following physical and chemical properties:

- (1) The reagent is readily and rapidly synthesized from F-18.
- (2) The reagent has adequate aqueous solubility in the neutral (4-8) pH range. By "adequate aqueous solubility" is meant that the reagent readily dissolves at up to a concentration comparable to a stoichiometric amount of the thiol-containing peptide used. If, for example, an antibody is being labeled, a typical antibody concentration is about 50mg/mL, which corresponds to a molar concentration of about 3×10^{-4} M. In this example, the reagent should be soluble at a concentration of about 3×10^{-4} M. With lower molecular weight peptide species, more peptide will dissolve without precipitation, and more reagent can be used. Because F-18 is carrier-free, lower concentrations of fluorination agents also might be effective.

- (3) The active halides of the reagent are not immediately hydrolyzed by water at neutral pH (pH 4-8). Thus, the halides should react more readily with SH or S⁻ than with H₂O. As long as the reagent is not immediately hydrolyzed by water (or by neutral buffer solutions), the selectivity and reactivity of the thiol group ensures an efficient peptide labeling reaction.

- (4) The leaving group X can be displaced rapidly, specifically, and near-quantitatively by free thiol moieties. A carbo-cationic center can be developed at the

carbon atom which is attacked by the nucleophile, for example, R_1 and R_2 can be electron-withdrawing groups. The presence of electron-withdrawing groups alpha to the -C-X functional group also facilitates fast displacement of the X moiety. Examples of useful electron-withdrawing groups include -COR', -CONR', -CO₂R', -COOH, -CONH₂, and -SO₃H, where R' is a C₁-C₆ alkyl or phenyl.

In addition, the presence of more than one leaving group in the labelling reagent can be advantageous. Multiple leaving groups, such as iodo groups, attached to the same carbon atom produce steric strain. When a reaction comprises the departure of a single leaving group, this steric strain is relieved, imparting faster reaction kinetics to the thiol displacement of the X group. Thus, in accordance with one embodiment of the invention, the labeling reagent comprises at least two leaving groups, such as two iodo groups.

In accordance with one embodiment of the present invention, the peptide is labeled with a labelling reagent of the general formula $^{18}\text{F}-(\text{CH}_2)_m-\text{CR}_1\text{R}_2-(\text{CH}_2)_n-\text{X}$, wherein n is 0, 1 or 2, m is 0, 1 or 2, and n+m is 0, 1, or 2, and X is a substitutable leaving group such as iodide, bromide, chloride, azide, tosylate, mesylate, nosylate, triflate, and the like. Alternatively, X is maleimide or a substituted maleimide, substituted, for example with one or two alkyl groups. Examples of suitable substituted maleimides include 3-methylmaleimide, 3,4-dimethylmaleimide and 3-sulfo-maleimide. R_1 and R_2 can be the same or different and, as discussed above, are chosen for the desirable physical properties they bring to the reagent. In general, R_1 and R_2 can be selected from the same groups as X, and can be the same as or different from X. Alternatively, R_1 and R_2 each independently can be hydrogen, a substituted or unsubstituted linear or branched alkyl group, or a carbonyl function such as an ester, amide or ketone, for example, -COOR', -CONR'₂, or COR', where R' is a C₁-C₆ alkyl or phenyl. Examples of suitable R_1 and R_2 groups or substituents thereon also include those which impart aqueous solubility, such as -CONH₂, carboxyl, hydroxyl, sulfonic acid and tertiary amine or quaternary ammonium.

Examples of suitable labelling reagents include $^{18}\text{F}-\text{Cl}_3$; $^{18}\text{F}-\text{CHI}_2$; $^{18}\text{F}-\text{Cl}_2\text{COOH}$; $^{18}\text{F}-\text{Cl}_2\text{COOCH}_3$; $^{18}\text{F}-\text{Cl}_2\text{CH}_2\text{OH}$; $^{18}\text{F}-\text{CHICH}_2\text{OH}$; $^{18}\text{F}-\text{CHICOOCH}_3$; $^{18}\text{F}-\text{Cl}_2\text{CH}_2\text{COOH}$; $^{18}\text{F}-\text{Cl}_2\text{CH}_2\text{N}^+(\text{CH}_3)_3$; $^{18}\text{F}-\text{Cl}_2\text{CH}_2\text{maleimide}$; $^{18}\text{F}-\text{Cl}_2\text{CONH}_2$; $^{18}\text{F}-\text{Cl}_2\text{CO}_2\text{CH}_3$; $^{18}\text{F}-\text{CHBr}_2$; $^{18}\text{F}-\text{CBr}_2\text{CH}_2\text{CH}_2-\text{SO}_3\text{H}$; $^{18}\text{F}-\text{CH}_2\text{Cl}_2\text{COOH}$; $^{18}\text{F}-\text{CH}_2\text{Cl}_2\text{CONH}_2$;

$^{18}\text{F-CHICO}_2\text{CH}_3$; $^{18}\text{F-Cl}_2\text{CONH}_2$; $^{18}\text{F-CHICONH}_2$; $^{18}\text{F-CBr}_2\text{CH}_2\text{OH}$; $\text{CF}_3\text{COCl}_2\text{-}^{18}\text{F}$; $\text{CH}_3\text{COCBr}_2\text{-}^{18}\text{F}$; $^{18}\text{F-CHBrCN}$; $^{18}\text{F-Cl}_2\text{CHCN}$; $\text{CBrF}_2\text{-}^{18}\text{F}$; $^{18}\text{F-CBr}(\text{CONH}_2)_2$, and $\text{C}_6\text{H}_5\text{-COCBr}_2\text{-}^{18}\text{F}$. Other suitable labeling reagents will be apparent to those skilled in the art.

5 The labeling reagent can be made by the F-18 fluorination of a corresponding compound. The following are examples of compounds which can be fluorinated to make the labeling reagents set forth above: Cl_4 ; CHI_3 ; $\text{CHI}_2\text{COOCH}_3$; Cl_3COOH ; $\text{Cl}_3\text{COOCH}_3$; $\text{Cl}_3\text{CH}_2\text{OH}$; $\text{CHI}_2\text{CH}_2\text{OH}$; $\text{Cl}_3\text{CH}_2\text{COOH}$; $\text{Cl}_3\text{CH}_2\text{N}^+(\text{CH}_3)_3$; $\text{Cl}_3\text{CH}_2\text{-maleimide}$; $\text{Cl}_3\text{-CONH}_2$; $\text{Cl}_3\text{-CO}_2\text{CH}_3$; CHIBr_2 ; $\text{CIBr}_2\text{CH}_2\text{CH}_2\text{-SO}_3\text{H}$; $\text{CH}_2\text{Cl}_3\text{COOH}$;
10 $\text{CH}_2\text{Cl}_3\text{CONH}_2$; $\text{CHI}_2\text{CO}_2\text{CH}_3$; Cl_3CONH_2 ; $\text{CHI}_2\text{CONH}_2$; $\text{CBr}_3\text{CH}_2\text{OH}$; CF_3COCl_3 ; $\text{CH}_3\text{COCBr}_3$; Br_2CHCN ; Cl_3CHCN ; CBr_2F_2 ; $\text{CBr}_2(\text{CONH}_2)_2$ and $\text{C}_6\text{H}_5\text{-COCBr}_3$. Other suitable compounds will be apparent to those skilled in the art.

 In accordance with another embodiment of the invention, the labeling reagent is an F-18 fluorinated alkene, wherein at least one of the two double-bonded carbon atoms
15 bears at least one leaving group selected from the group consisting of iodide, bromide, chloride, azide, tosylate, mesylate, nosylate and triflate. Examples of suitable fluorinated alkenes include $^{18}\text{F-CH=Cl}_2$, $^{18}\text{F-CI=CH}_2$, and $^{18}\text{F-CI=Cl}_2$. These labeling reagents can be made by the F-18 fluorination of corresponding compounds, such as ICH=Cl_2 ; $\text{Cl}_2=\text{CH}_2$; $\text{Cl}_2=\text{Cl}_2$. Other fluorinated alkenes useful in accordance with the
20 present invention will be apparent to those skilled in the art.

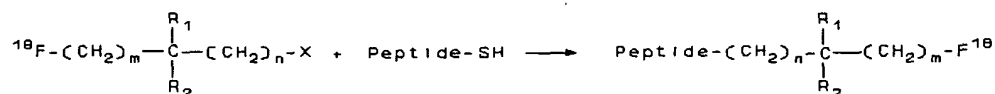
 F-18 can be obtained from cyclotrons after bombardment of O-18-enriched water with protons. The enriched water containing H- ^{18}F can be neutralized with a base having a counter-ion that is any alkali metal (M), such as potassium or another monovalent ion, and the water can be evaporated off to give a residue of M- ^{18}F , which
25 can be taken up in an organic solvent for further use. In general, the counter-ion is selected to enable the fluoride ion to react rapidly in an organic phase with a halogen. Potassium is generally used as a counter-ion because it is cheaper than cesium. However, with carrier-free F-18, trivial amounts of counter-ion are required, and counter-ion cost largely can be ignored.

30 Although potassium is useful as a counter-ion in accordance with the present invention, cesium is preferred to potassium because cesium is a larger ion with a more diffuse charge. Accordingly, cesium has looser ionic interactions with the small

fluoride atom, and therefore does not interfere with the nucleophilic properties of the fluoride ion. For similar reasons, potassium is preferred to sodium, and, in general, the suitability of a Ia metal as a counter-ion in accordance with the present invention increases as you go down the periodic table. Group Ib reagents, such as silver, also are
 5 useful as counter-ions in accordance with the present invention. Further, organic phase transfer-type ions, such as tetraalkylammonium salts, also can be used as counter-ions.

Because fluoride is the most electronegative element, it has a tendency to become hydrated and lose its nucleophilic character. To minimize this, the labeling reaction is preferably performed under anhydrous conditions. For example, fluoride (as
 10 potassium fluoride or as a complex with any of the other counter-ions discussed above) can be placed in organic solvents, such as acetonitrile or THF. With the help of agents which bind to the counter-ion, such as Kryptofix 2.2.2 (4,7,13,16,21,24-hexaoxa-1,10-diazabicyclo[8.8.8]-hexacosane), the fluoride ion is very nucleophilic in these solvents.

As discussed above, the labeling reagent is used to label targeting vectors
 15 comprising a thiol-containing peptide with F-18 according to the following reaction:



Alternatively, the labeling reagent is a F-18 fluorinated alkene, wherein at least one of the two double-bonded carbon atoms bears at least one leaving group selected from the group consisting of iodide, bromide, chloride, azide, tosylate, mesylate, nosylate and triflate. This F-18 fluorinated alkene labels targeting vectors in an analogous manner to
 20 the reaction set forth above.

Directing the reaction of the fluorinated labeling reagent towards free thiol groups on the targeting vector allows near-quantitative incorporation of F-18 into the targeting vector within a short time period. Generally, the reaction will be completed within a few minutes at room temperature, and complicated purification steps will not
 25 be necessary. Given the very short half-life of F-18, the speed of the reaction is very important. Moreover, because free F-18 exchanges readily with hydroxyl ions in hydroxyapatite crystals in bone, and, therefore, is a bone-seeking agent, the reduced

amount of free fluoride remaining in the final product also is an important advantage of the present invention.

5 The embodiments of the invention are further illustrated through examples which show aspects of the invention in detail. These examples illustrate specific elements of the invention and are not to be construed as limiting the scope thereof.

EXAMPLES

Fluorodiiodoacetic acid ($^{18}\text{F}-\text{Cl}_2\text{COOH}$)

10 100 mCi of F-18 fluoride (obtained from bombardment of O-18-enriched water) in dry tetrahydrofuran containing Kryptofix 2.2.2 (4,7,13,16,21,24-hexaoxa-1,10-diazabicyclo[8.8.8]hexacosane) and a slurry of potassium carbonate is treated with triiodoacetic acid. After a 30 minute reaction at room temperature, the desired labelling reagent, $^{18}\text{F}-\text{Cl}_2\text{COOH}$, is obtained and purified by reverse-phase column chromatography. This labelling reagent is then used to label a variety of thiol-
15 containing targeting vectors, or is shipped to clinical sites for the same usage.

F-18-Labeled Fab'-SH Fragment

A 1 mg vial of lyophilized Fab'-SH-NP4 (an anti-carcinoembryonic antigen antibody fragment) is reconstituted with 1 mL of a solution of $^{18}\text{F}-\text{Cl}_2\text{COOH}$ in 0.1 M sodium acetate buffer at pH 6. The reaction is allowed to proceed for 30 minutes at
20 room temperature.

An aliquot of the mixture is removed for analysis by HPLC using a size-exclusion sizing column and by ITLC (instant thin-layer chromatography) using silica gel-impregnated glass-fiber strips (Gelman Sciences). This analysis reveals that the antibody fragment's hinge-region thiol groups effect nucleophilic displacement of both
25 iodine atoms of $^{18}\text{F}-\text{Cl}_2\text{COOH}$, and that this reaction proceeds in near-quantitative yield. The F-18-labeled Fab' fragment is therefore ready for injection.

Fluorodiiodomethane ($^{18}\text{F}-\text{CHI}_2$)

A sample of 100 mCi of F-18 fluoride (obtained from bombardment of O-18-enriched water) in dry acetonitrile containing Kryptofix 222 and a slurry of potassium carbonate is treated with triiodomethane. After a 30 minute reaction at room
5 temperature the labelling reagent $^{18}\text{F}-\text{CHI}_2$ is obtained and purified by reverse-phase column chromatography. The labelling reagent is then used to label a variety of thiol-containing targeting vectors, or is shipped to clinical sites for the same usage.

F-18-Labeled Octreotide

A 1 mg vial of lyophilized, reduced octreotide (D-Phe-Cys-Phe-D-Trp-Lys-Thr-
10 Cys-Thr-ol) is reconstituted with 1 mL of a solution of $^{18}\text{F}-\text{CHI}_2$ (made up first in DMSO) in 0.1 M sodium acetate buffer at pH 6, containing 20% DMSO. The reaction is allowed to proceed for 30 minutes at room temperature. Alternatively, can be effected at elevated temperatures, and in non-aqueous solvents, e.g., DMSO, and later cooled and/or diluted for injection.

15 An aliquot of the labeling mixture is removed for analysis by HPLC using a size-exclusion sizing column and ITLC (instant thin-layer chromatography) using silica gel-impregnated glass-fiber strips (Gelman Sciences). This analysis reveals that the two cysteinyl thiol groups of octreotide effect the nucleophilic displacement of both iodo
atoms of $^{18}\text{F}-\text{CHI}_2$, and that this reaction proceeds in near-quantitative yield. The F-18-
20 labeled, recyclized (linkage: $-\text{S}-\text{CH}-^{18}\text{F}-\text{S}-$) octreotide peptide is therefore ready for injection.

Fluorodiiodoacetamide ($^{18}\text{F}-\text{Cl}_2\text{CONH}_2$)

100 mCi of F-18 fluoride (obtained from bombardment of O-18-enriched water) in dry tetrahydrofuran containing Kryptofix 2.2.2 (4,7,13,16,21,24-hexaoxa-1,10-
25 diazabicyclo[8.8.8]hexacosane) and a slurry of potassium carbonate is treated with triiodoacetamide. After a 30 minute reaction at room temperature, the desired labelling reagent, $^{18}\text{F}-\text{Cl}_2\text{CONH}_2$, is obtained and purified by reverse-phase column chromatography. This labelling reagent is then used to label a variety of thiol-containing targeting vectors, or is shipped to clinical sites for the same usage.

F-18-Labeled Cys-LHRH

5 A 1 mg vial of lyophilized Cys-LHRH (LHRH whose amine terminus bears an appended cysteine, in reduced, thiol, form) reconstituted with 1 mL of a solution of ^{18}F -CH₃CONH₂ in 0.1 M sodium acetate buffer at pH 6. The reaction is allowed to proceed for 2 hours at 50°C. The antibody modified peptide's thiol group effects nucleophilic displacement of the iodo atom of ^{18}F -I-CH₃CONH₂, and the reaction proceeds in near-quantitative yield. The F-18-labeled peptide is ready for injection.

10 It will be apparent to those skilled in the art that various modifications and variations can be made to this invention. Thus, it is intended that the present invention cover the modifications and variations of this invention provided they come within the scope of the claims and their equivalents.